

## User manual RTP<sup>®</sup> DNA/ RNA Virus Mini Kit

for simultaneous purification of viral DNA and RNA from human and animal serum and plasma samples, cell culture supernatants and other cell-free body fluids (e.g. urine), swabs, tissue biopsies and stool suspension



IVD

REF

1040100x0



STRATEC Molecular GmbH, D-13125 Berlin

## Instruction for the RTP<sup>®</sup> DNA/ RNA Virus Mini Kit

The **RTP<sup>®</sup> DNA/ RNA Virus Mini Kit** is a well established tool using RTP<sup>®</sup> technology for simultaneous isolation of high quality viral DNA and RNA from human and animal serum and plasma samples, cerebrospinal fluid, cell culture supernatants and other cell free body fluids, like urine, supernatant of stool suspension as well as from swabs or tissue biopsies for *in-vitro*-diagnostic purposes using a spin-filter format. Fresh or frozen plasma or serum from blood treated with anticoagulants like EDTA or citrate, *but not with heparin*. The customer convenient RTP<sup>®</sup> technology simplifies the process handling, reduces the handling steps with infectious material and allows a process monitoring.

Due to the high purity, the isolated viral DNA and RNA is ready to use for a broad panel of downstream applications or can be stored at – 80 °C for subsequent use.

The kit is neither suitable for isolation of viral DNA or RNA from whole blood or blood stains, nor for isolation of RNA or DNA from bacteria, fungi, plants.



Compliance with EU Directive 98/79/EC on *in vitro* medical devices.

Not for *in-vitro* diagnostic use in countries where the EU Directive 98/79/EC on *in vitro* medical devices is not recognized.

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The Invisorb<sup>®</sup> technology is covered by patents and patent applications: US 6,110363, US 6,043,354, US 6,037,465, EP 0880535, WO 9728171, WO 9534569, EP 0765335, DE 19506887, DE 10041825.2, WO 0034463.

Invisorb<sup>®</sup>, RTP<sup>®</sup> and InviMag<sup>®</sup> are registered trademarks of STRATEC Biomedical AG.

The PCR process is covered by US Patents 4,683,195, and 4,683,202 and foreign equivalents owned by Hoffmann-La Roche AG.

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## Kit contents of RTP® DNA/ RNA Virus Mini Kit

Store all kit components at room temperature (RT)!

	5 extractions	50 extractions	250 extractions
<b>Catalogue No.</b>	1040100100	1040100200	1040100300
<b>Extraction Tubes</b>	5	50	5 x 50
<b>Binding Solution</b> (fill with 99.7% Isopropanol)	3 x 1 ml (ready to use)	empty bottle (final volume 30 ml)	empty bottle (final volume 120 ml)
<b>Wash Buffer R1</b>	15 ml (ready to use)	20 ml (final volume 40 ml)	80 ml (final volume 160 ml)
<b>Wash Buffer R2</b>	15 ml (ready to use)	12 ml (final volume 60 ml)	50 ml (final volume 250 ml)
<b>Elution Buffer R</b>	2 ml	15 ml	30 ml
<b>RTA Spin Filter Set</b>	5	50	5 x 50
<b>RTA Receiver Tubes</b>	15	3 x 50	15 x 50
<b>Elution Tubes</b>	5	50	5 x 50
<b>Manual</b>	1	1	1
<b>Initial steps</b>	<b>Wash Buffer R1, Wash Buffer R2, Binding Solution</b> are ready to use	Fill 30 ml 99.7% <b>Isopropanol</b> (molecular biologic grade) into the empty bottle  Add 20 ml of 96 – 100 % ethanol to the bottle <b>Wash Buffer R1</b> , mix thoroughly and always keep the bottle firmly closed!  Add 48 ml of 96 – 100 % ethanol to the bottle <b>Wash Buffer R2</b> , mix thoroughly and always keep the bottle firmly closed!	Fill 120 ml 99.7% <b>Isopropanol</b> (molecular biologic grade) into the empty bottle  Add 80 ml of 96 – 100 % ethanol to the bottle <b>Wash Buffer R1</b> , mix thoroughly and always keep the bottle firmly closed  Add 200 ml of 96 – 100 % ethanol to the bottle <b>Wash Buffer R2</b> , mix thoroughly and always keep the bottle firmly closed!

## Symbols

	Manufacturer
	Lot number
	Catalogue number
	Expiry date
	Consult operating instructions
	Temperature limitation
	Do not reuse

## Storage

All buffers and kit components of the **RTP® DNA/ RNA Virus Mini Kit** including the **Extraction Tube** (incl. Lysis Buffer, Proteinase K, Carrier RNA and Internal Control DNA) should be stored well sealed and dry at room temperature (RT) and are stable for at least 12 months under these conditions.

**Wash Buffer R1 and R2** charged with ethanol should be appropriately sealed.

Before every use make sure that all components have room temperature. If there are any precipitates within the provided solutions solve these precipitates by warming up carefully.

**Room temperature is defined as range from 15 – 30°C.**

## Quality control and product warranty

STRATEC Molecular warrants the correct function of the **RTP® DNA/ RNA Virus Mini Kit** for applications as described in this manual. Purchaser must determine the suitability of the Product for its particular use. Should any Product fail to perform the applications as described in the manual, STRATEC Molecular will check the lot and if STRATEC Molecular investigates a problem in the lot, STRATEC Molecular will replace the Product free of charge.

STRATEC Molecular reserves the right to change, alter, or modify any Product to enhance its performance and design at any time.

In accordance with STRATEC Molecular's ISO 9001-2000 and ISO EN 13485 certified Quality Management System the performance of all components of the **RTP® DNA/ RNA Virus Mini Kit** have been tested separately against predetermined specifications routinely on lot-to-lot to ensure consistent product quality.

If you have any questions or problems regarding any aspects of **RTP® DNA/ RNA Virus Mini Kit** or other STRATEC Molecular products, please do not hesitate to contact us. A copy of STRATEC Molecular's terms and conditions can be obtained upon request or are presented at the STRATEC Molecular webpage.

**For technical support or further information please contact:**

**from Germany +49-(0)30-9489-2901/ 2910**

**from abroad +49-(0)30-9489-2907**

**or contact your local distributor.**

## Intended use

The **RTP® DNA/ RNA Virus Mini Kit** is the ideal tool for reliable and fast simultaneous isolation of high quality viral DNA and RNA from fresh or frozen human or mammalian serum, plasma, cerebrospinal fluid, cell culture supernatants and other cell free body fluids as well as from swabs or tissue biopsies.

For reproducible high yields an appropriate sample storage and quick operation under the rules for RNA and DNA operation is essential. The purified viral DNA and/or RNA is ready to use for *in vitro* diagnostic analysis only.

The isolation protocols as well as all buffers are optimized to provide high yield and purity of the extracted viral nucleic acids. The procedure requires minimal interaction by the user, allowing safe handling of potentially infectious samples.

THE PRODUCT IS INTENDED FOR USE BY PROFESSIONALS ONLY, SUCH AS TECHNICIANS, PHYSICIANS AND BIOLOGISTS TRAINED IN MOLECULAR BIOLOGICAL TECHNIQUES. It is designed to be used with any downstream application employing enzymatic amplification or other enzymatic modifications of DNA/ RNA followed by signal detection or amplification. Any diagnostic results generated by using the sample preparation procedure in conjunction with any downstream diagnostic assay should be interpreted with regard to other clinical or laboratory findings.

To minimize irregularities in diagnostic results, adequate controls for downstream applications should be used.

*The kit is in compliance with EU Directive 98/79/EC on in vitro medical devices. But it is not for in-vitro diagnostic use in countries where the EU Directive 98/79/EC on in vitro medical devices is not recognized.*

## Product use limitation

The kits are neither suitable for isolation of viral DNA or RNA from whole blood or blood stains, nor for isolation of RNA or DNA from bacteria, fungi, plants.

The included chemicals are only useable once.

Differing of starting material or flow trace may lead to inoperability; therefore neither a warranty nor guarantee in this case will be given, neither implied nor express.

The user is responsible to validate the performance of the STRATEC Molecular Product for any particular use. STRATEC Molecular does not provide for validation of performance characteristics of the Product with respect to specific applications. STRATEC Molecular Products may be used e.g. in clinical diagnostic laboratory systems conditioned upon the complete diagnostic system of the laboratory the laboratory has been validated pursuant to CLIA' 88 regulations in the U.S. or equivalents in other countries.

All Products sold by STRATEC Molecular are subject to extensive quality control procedures (according to ISO 9001-2000 and ISO EN 13485) and are warranted to perform as described herein. Any problems, incidents or defects shall be reported to STRATEC Molecular immediately upon detection thereof.

The chemicals and the plastic parts are for laboratory use only; they must be stored in the laboratory and must not be used for purposes other than intended.

The Product with its contents is unfit for consumption.

## Safety information

When and while working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles!

Avoid skin contact! Adhere to the legal requirements for working with biological material!

For more information, please consult the appropriate material safety data sheets (MSDS). These are available online in convenient and compact PDF format at [www.stratec.com](http://www.stratec.com) for each STRATEC Molecular Product and its components. If buffer bottles are damaged or leaking, WEAR GLOVES, AND PROTECTIVE GOGGLES when discarding the bottles in order to avoid any injuries.

STRATEC Molecular has not tested the liquid waste generated by the **RTP® DNA/ RNA Virus Mini Kit** procedure for residual infectious materials. Contamination of the liquid waste with residual infectious materials is highly unlikely, but cannot be excluded completely. Therefore, liquid waste must be considered infectious and be handled and discarded according to local safety regulations.

European Community risk and safety phrases for the components of the **RTP® DNA/ RNA Virus Mini Kit** to which they apply is listed are listed below as follows:

### Wash Buffer R1



warning

H302-312-332-412 EUH032 P273

### Extraction Tube L



danger

H302-314 EUH208 P260-280-305-351-338-310

<b>H302:</b>	Harmful if swallowed.
<b>H312:</b>	Harmful in contact with skin.
<b>H332:</b>	Harmful if inhaled.
<b>H412:</b>	Harmful to aquatic life with long lasting effects.
<b>EUH032:</b>	Contact with acids liberates very toxic gas.
<b>H314:</b>	Causes severe skin burns and eye damage.
<b>EUH208:</b>	Contains Proteinase K. May produce an allergic reaction.
<b>P305+P351+P338:</b>	IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
<b>P273:</b>	Avoid release to the environment.
<b>P260:</b>	Do not breathe dust/fume/gas/mist/vapours/spray.
<b>P280:</b>	Wear protective gloves/protective clothing/eye protection/face protection.
<b>P310:</b>	Immediately call a POISON CENTER or doctor/physician

**Emergency medical information can be obtained 24 hours a day from infotrac:**

outside of USA: 1 – 352 – 323 – 3500  
in USA : 1 – 800 – 535 – 5053

## Product characteristic of the RTP<sup>®</sup> DNA/ RNA Virus Mini Kit

Starting material	Yield	Time
up to 200 µl cell free body fluids; swab material, cell culture supernatants  up to 400 µl rinse liquid from swab  1 x 10 <sup>6</sup> mammalian cells max. 10 mg tissue sample 100 µl/ 100 mg stool sample	depends on the sample (storage and source)  <b>Note:</b> The added Carrier RNA will account for most of the eluted RNA. Quantitative RT-PCR is recommended for determination of the viral RNA or DNA yield.	20 minutes

The **RTP<sup>®</sup> DNA/ RNA Virus Mini Kit** provides a fast and efficient way for reliable simultaneous isolation of high quality viral DNA and RNA from a diverse range of starting material. The procedures are suitable for use with plasma or serum; either can contain citrate or EDTA (no heparin) and other samples. Samples can be fresh, lyophilized or frozen, provided they have not been frozen and thawed more than ones.

The amount of purified viral DNA and RNA in the **RTP<sup>®</sup> DNA/ RNA Virus Mini Kit** procedure depend on the sample type, the virus titer, sample source, transport, storage, and age.

The kits use the patented RTP<sup>®</sup> technology, whose special feature is an **Extraction Tube** containing preformulated solid lysis reagent, Proteinase K and Carrier Nucleic Acids. The technology allows the reduction of reagent preparation steps and of handling steps with infectious material.

Using the **RTP<sup>®</sup> DNA/ RNA Virus Mini Kit**, all types of samples are transferred into the Extraction Tubes together with ddH<sub>2</sub>O to adjust a final sample volume of 400 µl. The prefilled buffer and enzymes lyse the samples, stabilize the viral nucleic acids and enhance the selective viral RNA and DNA adsorption to the membrane in the RTA Spin Filter. Contaminants are removed by repeated wash steps and the purified viral DNA and/ or RNA can be eluted in a small volume of **Elution Buffer R**.

The advantage of the kit results from the simultaneous isolation of nucleic acids from DNA and RNA viruses. That enables the user to test the sample for all kinds of RNA and DNA viruses which are of interest after one preparation.

Yield and quality of isolated viral DNA and RNA is suitable for any molecular-diagnostic detection system. The diagnostic tests should be performed according to manufacturer's specifications. Due to the high purity, the isolated viral DNA/ RNA is ready to use for a broad panel of downstream applications or can be stored at –80°C for subsequent use.

- (RT)-PCR
- Real-time PCR (quantitative (RT)-PCR, like TaqMan und LightCycler technology)
- cDNA syntheses
- Microarray-application



The purification procedure is rapid and does not require a phenol/ chloroform extraction or  $\beta$ -Mercaptoethanol. Only a minimum of manual work by the user is necessary. The procedure is designed to avoid sample-to-sample cross-contaminations and allow safe handling of potentially infectious samples. The procedure is highly suited for simultaneous processing of multiple samples. Traditional time-killing procedures can be replaced using the **RTP® DNA/ RNA Virus Mini Kit**. All kit components can be stored at room temperature.

STRATEC Molecular also offers in combination with magnetic bead the **InviMag® Virus DNA/ RNA Mini Kit/ KFml**.

**Note:** *Systems isolating simultaneously DNA and RNA from viruses using buffers adapted for the binding of DNA and RNA, but the optimal binding conditions of RNA and DNA are different, so that such solutions can show a little reduction in sensitivity in comparison to kits optimized to one kind of nucleic acid isolation.*

STRATEC Molecular offers also kits for the separate isolation of viral DNA or viral RNA from serum and plasma in single tube or 96 well format, **Invisorb® Spin Virus RNA Mini Kit**, the **Invisorb® Spin Virus DNA Mini Kit**. The **Invisorb® Virus RNA HTS 96-Kit/ X** is designed for use on X-tractor Gene™ (Corbett Robotics) and the the **Invisorb® Virus RNA Mini Kit KF96** for use on KingFisher 96.

If you are interested in using the kit on a laboratory workstation, please do not hesitate to contact our technical support (+49 (0)30-9489-2907). For technical support or further information please contact: **+49 - (0)30-9489-2901/ 2907/ 2910 or your local distributor.**

## **Internal controls/ Extraction control**

Internal Controls (IC) from the PCR assay provider can be used as extraction controls if the fragments are longer than 100 bp. In this case they have to be added after finalization of the lysis step.

**Attention:** don't add directly these Internal Controls to the sample!

## Principle

The **RTP® DNA/ RNA Virus Mini Kits** procedure comprises following steps:

1. lysis of the virus particles
2. adjustment of the binding conditions
3. binding the viral nucleic acids in the lysate to the membrane of a Spin Filter RTA
4. washing of the membrane and elimination of contaminants and ethanol
5. elution of the viral nucleic acids

This manual contains 4 protocols.

## Sampling and storage of starting material

Best results are obtained using freshly extracted samples. As long as the samples are not shock frosted with liquid nitrogen or are incubated with RNase inhibitors or denaturing reagents, the viral RNA is not secured. Therefore it is essential, that samples are immediately flash frozen subsequent to the harvesting by using liquid nitrogen and are stored at -80°C. Viral RNA contained in such deep frozen samples is stable for months. Viral RNA purification should be processed as soon as possible. Samples can also be stored in the dissolved Lysis Buffer in the Extraction Tubel or II for 1 h at room temperature, overnight at 4°C, and for long term storage at -80°C. Storage under deep frozen conditions is recommended.

Serum, plasma, urine, cerebrospinal fluid or other cell free body fluids, as well as cell culture supernatants, swabs, and stool samples can be stored on ice for 1 - 2 hours, for short time (up to 24 h) samples may be stored at -20°C. For long term storage, we recommend freezing samples at -80°C. Multiple thawing and freezing before isolating the viral RNA should be avoided.

### **Serum and plasma (and other cell free body fluids)**

Following centrifugation, plasma or serum from blood treated with anticoagulants like EDTA or citrate, *but not with heparin*, can be stored at 2–8°C for up to 6 hours. For long-term storage, freezing at -20°C to -80°C in aliquots is recommended. Repeated freezing and thawing cycles must be avoided because denaturation and precipitation of proteins result in a decrease of the virus titer and thereby reduce the yield of the extracted viral RNA. Occurring cryoprecipitates can be pelleted by briefly centrifuging (6.800 x g for 3 minutes). The cleared supernatant should be removed, without disturbing the pellet, and processed immediately. This step will not reduce viral titers.

### **Tissue samples (biopsy material or frozen section)**

Best results are obtained with fresh material or material that has been immediately flash frozen and stored at -20°C or -80°C. Repeated freezing and thawing of stored samples should be avoided, since this leads to reduced RNA yield. Use of poor quality starting material influences the RNA yield, too. The amount of purified RNA in the **RTP® DNA/ RNA Virus Mini Kit** procedure using up to 20 mg tissue sample, depends on kind of starting material. The thawing process could be proceed, e.g. directly in the **Extraction Tubes**.

### **Cell culture supernatants**

Best results are obtained with fresh material or material that has been immediately frozen and stored at -20°C or -80°C after winning of the cell culture supernatant. Repeated freezing and thawing of stored samples can influence the sensitivity.

### **Swabs:**

The protocol works with fresh prepared swabs or rinsed liquid from swabs or mouth brushes. Best results are obtained with fresh material or material that has been immediately frozen and stored at -20°C or -80°C.

STRATEC Molecular will be released of its responsibilities if other sample materials than described in the Intended Use are processed or if the sample preparation protocols are changed or modified.

## Procedure

### Lysis

Samples are lysed under denaturing conditions at elevated temperatures. Due to the strong denaturing lyses conditions in the presence of **Proteinase K** and the **Lysis Buffer** cells are quickly broken and RNases and DNases are inactivated simultaneously. The viral RNA and DNA are secured. The addition of **Carrier RNA** (provided in the Extraction Tube) is necessary for the enhancement of viral DNA/ RNA recovery so a very small number of viral DNA/ RNA molecules will also be purified. **Carrier RNA** also stabilizes nucleic acids in samples with very small nucleic acid concentrations.

### Binding viral RNA

After adding **Binding Solution** to optimize the binding of viral DNA/ RNA to the RTA Spin Filter membrane, the lysate will be applied onto the RTA Spin Filter and the viral DNA/ RNA are bound to the surface of the RTA Filter membrane as the lysate is drawn through by centrifugation.

### Removing residual contaminants

Contaminants are efficiently washed away using **Wash Buffer R1** and **R2**, while the viral DNA/ RNA remain bound to the membrane of the RTA Spin Filter.

### Elution

High quality viral DNA/ RNA is eluted from the membrane using **Elution Buffer R** or RNase free water. The eluted DNA/ RNA is ready to use in different subsequent applications.

## Important notes

### Important points before starting a protocol

Immediately upon receipt of the Product, inspect the Product and its components as well as the package for any apparent damages, correct quantities and quality. If there are any unconformities you have to notify STRATEC Molecular in writing with immediate effect upon inspection thereof. If buffer bottles are damaged, contact the STRATEC Molecular Technical Services or your local distributor. In case of liquid spillage, refer to "Safety Information" (see page 7). Do not use damaged kit components, since their use may lead to poor kit performance.

1. Always change pipet tips between liquid transfers. To avoid cross-contaminations, we recommend the use of aerosol-barrier pipet tips.
2. All centrifugation steps are carried out at room temperature.
3. When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles.
4. Discard gloves if they become contaminated.
5. Do not combine components of different kits unless the lot numbers are identical.
6. Avoid microbial contamination of the kit reagents.
7. To minimize the risk of infections from potentially infectious material, we recommend working under laminar air-flow until the samples are lysed.
8. This kit should only be used by trained personnel.

## Important indications

### Preparing viral RNA

When preparing viral RNA, work quickly during the manual steps of the procedure.

The Lysis Buffer in the Extraction Tube simplifies viral RNA isolation by combining efficient lysis of the starting material and the inactivation of exogenous and endogenous RNases. Special care should be taken to avoid contaminations with RNases when handling Elution Buffer R.

### Storing samples

Frozen serum or plasma samples must not be thawed more than once. Repeated freeze –thawing leads to denaturation and precipitation of proteins, resulting in reduced viral titers and therefore reduced yields of viral nucleic acids. In addition cryoprecipitate formed during freeze –thawing will clog the RTA Spin Filter membrane.

### Carrier RNA

Carrier RNA serves two purposes. Firstly, it enhances the binding of viral acids to the RTA Spin Filter membrane, especially if there are very few target molecules in the sample. Secondly, the addition of large amounts of Carrier RNA reduces the chance of viral nucleic acid degradation in the rare event that RNase or DNase molecules are not denaturated by the salts and detergents in the Lysis Buffer in the Extraction Tube.

### Internal Control

The use of an internal control is recommended when using the **RTP® Virus DNA/ RNA Mini Kit** in combination with diagnostic amplification systems.

### Eluting viral RNA and DNA

For downstream applications, that require small starting volumes, elution volume may be reduced to 40 µl Elution Buffer R.

The volume of eluate recovered may be up to 5 µl less than the volume of elution buffer applied to the RTA Spin Filter. The volume of eluate recovered depends on the nature of the sample.

### Handling of RTA Spin Filter

Due to the sensitivity of viral DNA/ RNA amplification technologies the following precautions are necessary when handling RTA Spin Filter to avoid cross-contamination between sample preparations.

1. carefully apply the sample or solution to the RTA Spin Filter, pipet the sample into the filter without wetting the rim of the column
2. always change pipet tips between liquid transfers, we recommend the use of aerosol barrier pipet tips
3. avoid touching the RTA Spin Filter membrane with the pipet tip

### Yield and quality of viral DNA/ RNA

Different amplification systems vary in efficiency depending on the total amount of nucleic acid present in the reaction. Eluates derived by this kit will contain Carrier-RNA, which will greatly exceed the amount of the isolated NA.

Yields of viral nucleic acids isolated from biological samples are usually low concentrated and therefore almost impossible to determine photometrically.\*

\*. Keep in mind that the Carrier-RNA (5 µg per 200 µl sample) will account for most of the present RNA.

The kit is suitable for downstream analysis with NAT techniques, for examples qPCR, RT qPCR, LAMP, LCR. Diagnostic assays should be performed according to the manufacturer's instructions.

Quantitative RT-PCR is recommended for determination of viral RNA yield.

*\* In Gel Electrophoresis and in Capillary Electrophoresis, RNA extracted with the provided kit looks like degraded cause the kit contains Carrier RNA, this is poly A RNA in fragments of 100 up to 1000 bases. The kit is not dedicated for applications using this kind of analysis.*

## Preparing buffers

<b>5 viral DNA/ RNA-extractions:</b>
Binding Solution, Wash Buffer R1 and R2 are ready to use!
<b>50 viral DNA/ RNA-extractions:</b>
Fill 30 ml 99.7% Isopropanol (molecular biologic grade) into the empty bottle Add 20 ml 96-100% ethanol to the bottle <b>Wash Buffer R1</b> . Add 48 ml 96-100% ethanol to each bottle <b>Wash Buffer R2</b> .
<b>250 viral DNA/ RNA-extractions:</b>
Fill 120 ml 99.7% Isopropanol (molecular biologic grade) into the empty bottle Add 80 ml 96-100% ethanol to each bottle <b>Wash Buffer R1</b> . Add 200 ml 96-100% ethanol to each bottle <b>Wash Buffer R2</b> .

## Harvesting $1 \times 10^6$ mammalian cells

<b>Cells grown in suspension:</b>	Spin up to $1 \times 10^6$ cells for 5 min at $240 \times g$ (1.500 rpm). Discard the supernatant and remove all media completely.
<b>Cells grown in a monolayer:</b>	In large culture vessels (dishes > 100 cm <sup>2</sup> ) detach cells by trypsinization. Transfer the cells to a centrifuge tube and sediment by centrifugation at $240 \times g$ (1.500 rpm) for 5 minutes. Remove the supernatant completely. In small culture vessels (96-, 24-, 12-, 6-well plates, flasks) discard the media completely and continue with the lysis immediately.

**Important:** Some specific cell culture media may inhibit lysis efficiency or downstream reactions.

For special comments on the diverse **sample preparation procedures** see the respective protocol on page 16.

## Equipment and reagents to be supplied by user

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information's, please consult the appropriate material safety data sheets (MSDS). (See our webpage: [www.strattec.com](http://www.strattec.com))

- Microcentrifuge  $\geq 11.000 \times g$  ( $\geq 11.000$  rpm)
- Thermomixer (65°C - 95°C)
- ddH<sub>2</sub>O
- Ethanol (96-100%)
- 2.0 ml reaction tubes (optional)
- Measuring cylinder (250 ml)
- Disposable gloves
- Pipet with tips
- 1.5 ml reaction tubes
- Isopropanol \*

\*The **RTP® Virus DNA/ RNA Mini Kit** is validated with 2-Propanol; Rotipuran >99.7%, p.a., ACS, ISO (Order no. 6752) from **Carl Roth**

\* Possible suppliers for Isopropanol:


**Carl Roth**  
2-Propanol  
Rotipuran >99.7%, p.a., ACS, ISO  
Order no. 6752

**Applichem**  
2-Propanol für die Molekularbiologie  
Order no. A3928

**Fa. Sigma**  
2-Propanol  
Order no. 59304-1L-F

## Scheme of the RTP<sup>®</sup> DNA/ RNA Virus Mini Kit

Please work quickly and perform all extraction steps at room temperature (RT)!

	<p><b>Please read the protocols carefully prior to the start of the preparation procedure!</b></p> <p>transfer 200 µl sample into the provided <b>Extraction Tubes</b>  add 200 µl <b>dd H<sub>2</sub>O</b>  for samples which have a smaller volume than 200 µl please fill up to a total volume of 400 µl with ddH<sub>2</sub>O  incubate for 15 minutes at 65°C in a thermomixer  incubate for 10 minutes at 95°C in a thermomixer (optional)</p> <p>for optimal binding conditions add 400 µl <b>Binding Solution</b> and mix the sample completely by pipetting up and down or by vortexing</p> <p>transfer the sample on the RTA Spin Filter  incubate for 1 min  centrifuge for 2 min at 11.000 x g (11.000 rpm)  discard the flow-through with the RTA Receiver Tube  put the RTA Spin Filter in a new RTA Receiver Tube</p> <p>pipet 500 µl <b>Wash Buffer R1</b> onto the RTA Spin Filter  centrifuge for 1 min at 11.000 x g (11.000 rpm)  discard the flow-through and the RTA Receiver Tube  transfer the RTA Spin Filter into a new RTA Receiver Tube</p> <p>pipet 700 µl <b>Wash Buffer R2</b> onto the RTA Spin Filter  centrifuge for 1 min at 11.000 x g (11.000 rpm)  discard the flow-through and the RTA Receiver Tube  transfer the RTA Spin Filter into a new RTA Receiver Tube</p> <p>to eliminate any traces of ethanol, centrifuge again for 4 min at maximum speed  discard the RTA Receiver Tube</p> <p>transfer the RTA Spin Filter into a RNase free 1.5 ml Elution Tube  pipet 60 µl of <b>Elution Buffer R</b> (<u>preheated to 65°C</u>) directly onto the membrane of the RTA Spin Filter,  incubate for 3 min  centrifuge for 1 min at 11.000 x g (11.000 rpm)  discard the RTA Spin Filter and place the eluted viral DNA/ RNA immediately on ice</p>
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## Instructions - The following notes are valid for all protocols:

- Note:** The DNA/ RNA can also be eluted with a lower (but not lower than 40 µl) or a higher volume of Elution Buffer R (depends on the expected yield or needed concentration of the DNA/ RNA).  
The eluate contains viral DNA and/ or viral RNA.  
If the starting material is e.g. tissue the eluate contains further genomic DNA.
- Important** After extraction place the Elution Tube on ice. For long time storage place the nucleic acids at –20°C or –80°C.
- Note:** The centrifugation steps were made with the **Centrifuge 5415 D from Eppendorf**.  
The indicated **rpm amounts** are referring to this centrifuge.

### Protocol 1: Simultaneous isolation of total nucleic acids (DNA and RNA) from cell free body fluids

Please read the protocols carefully prior to the start of the preparation procedure!

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**Important Note:** Prewarm the needed amount of **Elution Buffer R** to 65°C for the final elution step. The protocol has been optimized for the isolation of total nucleic acids from body fluids of 200 µl. For samples which have a smaller volume than 200 µl please fill up to a total volume of 400 µl with ddH<sub>2</sub>O.

#### 1. Sample Lysis

Mix 200 µl of the sample with 200 µl of ddH<sub>2</sub>O.  
Transfer the sample into the provided Extraction Tube. Close the cap and vortex shortly.  
Place the Extraction Tubes into a thermomixer and incubate under continuously shaking for 15 minutes at 65°C and for 10 minutes at 95°C, this leads to higher sensitivity.

#### 2. Binding of the DNA and RNA

Add 400 µl **Binding Solution** to the provided **Extraction Tube** and mix the sample completely by pipetting up and down or by vortexing. Transfer the sample into the RTA Spin Filter Set. Close the cap and centrifuge for 2 minutes at 11.000 x g (11.000 rpm).  
Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

#### 3. First Washing of the RTA Spin Filter

Add 500 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.  
Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

#### 4. Second Washing of the RTA Spin Filter

Add 700 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.  
Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.  
Remove the residual ethanol by final centrifugation for 4 minutes at maximum speed.  
Discard the RTA Receiver Tube with filtrate.

#### 5. Elution of the DNA/ RNA

Place the Spin Filter into a 1.5 ml Elution Tube and add 60 µl of the **Elution Buffer R** (prewarmed to 65°C) directly onto the RTA Spin Filter surface.  
Incubate for 3 minutes at RT and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.

## **Protocol 2: Simultaneous isolation of total nucleic acids (DNA and RNA) from swab material**

**Please read the protocols carefully prior to the start of the preparation procedure!**

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**Important Note:** Prewarm the needed amount of **Elution Buffer R** to 65°C for the final elution step.

### **1. Sample Lysis**

Place the swab into the provided Extraction Tube and add 400 µl of ddH<sub>2</sub>O. Place the Extraction Tube into a thermomixer and incubate under continuously shaking for 15 minutes at 65°C and for 10 minutes at 95°C, which leads to higher sensitivity.

**Important Note:** To get maximum yield of viral nucleic acids it is essential to leave the swab during the complete lysis time into the reaction tube. It is possible to cut the shaft of the swab, so that you can close the cap of the Extraction Tube (II). It is also possible to do the lysis step with opened cap. The removing of the swab from the Extraction Tube (II) ahead of time will lead to a dramatically reduced final yield!  
After lysis time carefully squeeze out the swab on the wall of the tube and discard the swab

### **2. Binding of the DNA and RNA**

Add 400 µl **Binding Solution** to the provided **Extraction Tube** and mix the sample completely by pipetting up and down or by vortexing. Transfer the sample into the RTA Spin Filter Set. Close the cap and centrifuge for 2 minutes at 11.000 x g (11.000 rpm). Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

### **3. First Washing of the RTA Spin Filter**

Add 500 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuges at 11.000 x g (11.000 rpm) for 1 minute.

Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

### **4. Second Washing of the RTA Spin Filter**

Add 700 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.

Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

Remove the residual ethanol by final centrifugation for 4 minutes at maximum speed. Discard the RTA Receiver Tube with filtrate.

### **5. Elution of the DNA/ RNA**

Place the RTA Spin Filter into a 1.5 ml Elution Tube. Add 60 µl of the **Elution Buffer R** (prewarmed to 65°C) directly onto the RTA Spin Filter surface. Incubate for 3 minutes at RT and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.



### **Protocol 3: Simultaneous isolation of total nucleic acids (DNA and RNA) from cell culture supernatants**

**Please read the protocols carefully prior to the start of the preparation procedure!**

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**Important Note:** Prewarm the needed amount of **Elution Buffer R** to 65°C for the final elution step.

#### **1. Sample Lysis**

Mix 200 µl of the cell culture supernatant (cell culture media) with 200 µl of ddH<sub>2</sub>O. Transfer the sample into the provided Extraction Tube. Close the cap and vortex shortly. Place the Extraction Tube into a thermomixer and incubate under continuously shaking for 15 minutes at 65°C and for 10 minutes at 95°C, which leads to higher sensitivity.

#### **2. Binding of the DNA and RNA**

Add 400 µl **Binding Solution** to the provided **Extraction Tube** and mix the sample completely by pipetting up and down or by vortexing. Transfer the sample into the Spin Filter Set. Close the cap and centrifuge for 2 minutes at 11.000 x g (11.000 rpm). Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

#### **3. First Washing of the RTA Spin Filter**

Add 500 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuges at 11.000 x g (11.000 rpm) for 1 minute. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

#### **4. Second Washing of the RTA Spin Filter**

Add 700 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge at 11.000 x g (11.000 rpm). for 1 minute. Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube. Remove the residual ethanol by final centrifugation for 4 minutes at maximum speed. Discard the RTA Receiver Tube with filtrate.

#### **5. Elution of the DNA/ RNA**

Place the RTA Spin Filter into a 1.5 ml Elution Tube. Add 60 µl of the **Elution Buffer R** (prewarmed to 65°C) directly onto the RTA Spin Filter surface. Incubate for 3 minutes at RT and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.

## **Protocol 4: Simultaneous isolation of total nucleic acids (DNA and RNA) from tissue biopsies**

**Please read the protocols carefully prior to the start of the preparation procedure!**

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**Important Note:** Prewarm the needed amount of **Elution Buffer R** to 65°C for the final elution step.

### **1. Sample Lysis**

Transfer 1 - 10 mg of the tissue sample into the provided Extraction Tube. Add 400 µl of ddH<sub>2</sub>O. Close the cap and vortex shortly. Place the provided Extraction Tube into a thermomixer and incubate under continuously shaking for 15 minutes at 65°C and for 10 minutes at 95°C, which leads to higher sensitivity.  
Lysis time at 65°C can be increased up to 30 minutes.

**Important:** A longer lysis time could reduce the final yield and the quality of some viral RNA species.

After lysis centrifuge the sample at max. speed for 1 minute to spin down unlysed material. Transfer the cleared supernatant completely into a 1.5 ml reaction tube (not provided).

### **2. Binding of the DNA and RNA**

Add 400 µl **Binding Solution** to the 1.5 ml reaction tube and mix the sample completely by pipetting up and down or by vortexing. Transfer the sample into the RTA Spin Filter Set. Close the cap and centrifuge for 2 minutes at 11.000 x g (11.000 rpm). Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

### **3. First Washing of the RTA Spin Filter**

Add 500 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuges at 11.000 x g (11.000 rpm). for 1 minute.  
Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

### **4. Second Washing of the RTA Spin Filter**

Add 700 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.  
Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.  
Remove the residual ethanol by final centrifugation for 4 minutes at maximum speed.  
Discard the RTA Receiver Tube with filtrate.

### **5. Elution of the DNA/ RNA**

Place the RTA Spin Filter into a 1.5 ml Elution Tube. Add 60 µl of the **Elution Buffer R** (prewarmed to 65°C) directly onto the RTA Spin Filter surface. Incubate for 3 minutes at RT and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.

## **Protocol 5: Extraction of viral nucleic acids from supernatant of stool suspension**

**Please read the protocols carefully prior to the start of the preparation procedure!**

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**Important Note:** Prewarm the needed amount of **Elution Buffer R** to 65°C for the final elution step.

### **1. Sample Lysis**

Transfer 100 µl/ 100 mg stool sample into a 2 ml tube and add 900 µl RNase-free Water. Vortex the sample for 30 s followed by a 1 min centrifugation step at 12.000 x g (13.000 rpm).

Transfer 400 µl virus containing supernatant into the Extraction Tube (prevent the aspiration of swimming particles).

Place the provided Extraction Tube into a thermomixer and incubate under continuously shaking for 15 minutes at 65°C and for 10 minutes at 95°C, which leads to higher sensitivity.

*If you want to add nucleic acids for extraction control, please add at this point*

**Important:** A longer lysis time could reduce the final yield and the quality of some viral RNA species.

### **2. Binding of the DNA and RNA**

Add 400 µl **Binding Solution** to the 1.5 ml reaction tube and mix the sample completely by pipetting up and down or by vortexing. Transfer the sample into the RTA Spin Filter Set. Close the cap and centrifuge for 2 minutes at 11.000 x g (11.000 rpm). Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

### **3. First Washing of the RTA Spin Filter**

Add 500 µl **Wash Buffer R1** to the RTA Spin Filter and centrifuges at 11.000 x g (11.000 rpm). for 1 minute.

Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

### **4. Second Washing of the RTA Spin Filter**

Add 700 µl **Wash Buffer R2** to the RTA Spin Filter and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.

Discard the RTA Receiver Tube with filtrate and place the RTA Spin Filter into a new RTA Receiver Tube.

Remove the residual ethanol by final centrifugation for 4 minutes at maximum speed.

Discard the RTA Receiver Tube with filtrate.

### **5. Elution of the DNA/ RNA**

Place the RTA Spin Filter into a 1.5 ml Elution Tube. Add 60 µl of the **Elution Buffer R** (prewarmed to 65°C) directly onto the RTA Spin Filter surface. Incubate for 3 minutes at RT and centrifuge at 11.000 x g (11.000 rpm) for 1 minute.

## Troubleshooting

Problem	Probable cause	Comments and suggestions
<b>clogged RTA Spin-Filter</b>	insufficient lysis, homogenization and/or too much starting material	increase lysis time increase g-force and/ or centrifugation time reduce amount of starting material all centrifugation steps should be conducted at room
<b>low amount of extracted DNA/ RNA</b>	insufficient lysis  incomplete elution      insufficient mixing of the sample with <b>Binding Solution</b>   incomplete removal of cell culture medium	increase lysis time  reduce amount of starting material. Overloading of RTA Spin Filter reduces yield  prolong the incubation time with prewarmed Elution Buffer R to 5 -10 min. Do the elution steps twice Take higher volume of <b>Elution Buffer R</b> .  mix sample sufficient by pipetting up and down with <b>Binding Solution</b> prior to transfer the sample onto the Spin Filter RTA  make sure that the cell culture medium is complete removed after the cell harvest
<b>low concentration of extracted DNA/ RNA</b>	too much <b>Elution Buffer</b>  incorrect storage of starting material	elute the DNA/ RNA twice with lower volume of <b>Elution Buffer R</b>  ensure that the storage of starting material was correct avoid thawing of the material
<b>DNA/ RNA does not perform well in downstream- applications (e.g. RT- PCR or PCR)</b>	ethanol carryover during elution  salt carryover during elution	increase g-force or centrifugation time when drying the RTA Spin Filter  ensure that the Wash Buffers are at room temperature check up the <b>Wash Buffers</b> for salt precipitates. If there are any precipitates, dissolve these precipitates by carefully warming

## Appendix

### General notes on handling DNA

#### Nature of DNA

The length and delicate physical nature of DNA require careful handling to avoid damage due to shearing and enzymatic degradation. Other conditions that affect the integrity and stability of DNA include acidic and alkaline environments, high temperature, and UV irradiation. Careful isolation and handling of high molecular weight DNA is necessary to ensure it will work well in various downstream applications. Damaged DNA could perform poorly in applications such as genomic Southern blotting, long-template PCR, and construction of cosmid libraries.

#### Handling fresh and stored material before the extraction of DNA

For the isolation of genomic DNA from cells or tissues, use either fresh samples or samples that have been quickly frozen in liquid nitrogen and stored at -70°C. This procedure minimizes degradation of crude DNA by limiting the activity of endogenous nucleases.

#### Storage of DNA

Store genomic DNA at +2 to +8°C. Storing genomic DNA at -15 to -25°C can cause shearing of DNA, particularly if the DNA is exposed to repeated freeze-thaw cycles. Plasmid DNA and other small circular DNAs can be stored at +2 to +8°C or at -15 to -25°C.

#### Drying, dissolving and pipetting DNA

Avoid overdrying genomic DNA after ethanol precipitation. It is better to let it air dry than to use a vacuum, although vacuum drying can be used with caution. Plasmid DNA and other small circular DNAs can be vacuum-dried.

To help dissolve the DNA, carefully invert the tubes several times after adding buffer and tap the tube gently on the side. Alternatively let the DNA stand in buffer overnight at +2 to +8°C. Minimize vortexing of genomic DNA since this can cause shearing.

Avoid vigorous pipetting. Pipetting genomic DNA through small tip openings causes shearing or nicking. One way to decrease shearing of genomic DNA is to use special tips that have wide openings designed for pipetting genomic DNA. Regular pipette tips pose no problem for plasmid DNA and other small.

#### Quantification

Quantification of DNA and RNA from this assay must be done by means of qPCR or Reverse Transcriptase qPCR. All other methods will be disturbed by the included Carrier Nucleic Acids as well as DNA or RNA which is co purified.

## General notes on handling RNA

RNA is far less stable than DNA. It is very sensitive to degradation by endogenous RNases in the biological material and exogenous RNases which are permanently present everywhere in the lab. To achieve satisfactory qualitative and quantitative results in RNA preparations, contaminations with exogenous RNases has to be reduced as much as possible. Avoid handling bacterial cultures, cell cultures, or other biological sources of RNases in the same lab where the RNA purification is to be carried out.

All glassware should be treated before use to ensure that it is RNase free. Glassware should be cleaned with detergent, thoroughly rinsed and oven baked at 240°C for four or more hours before use. Autoclaving alone will not completely inactivate many RNases. Oven baking will both inactivate RNases and ensure that no other nucleic acids (such as Plasmid DNA) are present on the surface of the glassware. You can also clean glassware with 0.1% DEPC (diethyl pyrocarbonate). The glassware must stand 12 hours at 37°C and then be autoclaved or heated to 100°C for 15 minutes to remove residual DEPC.

- **Electrophoresis tanks** should be cleaned with detergent solution (e.g. 0.5% SDS), thoroughly rinsed with RNase free water, and then rinsed with ethanol and allowed to dry.
- **Non-disposable plasticware** should be treated before use to ensure that it is RNase free. plastic ware should be thoroughly rinsed with 0.1 M NaOH, 1 mM EDTA followed by RNase free water. You can also take chloroform-resistant plastic ware rinsed with chloroform to inactivate RNases.
- All **buffers** must be prepared from DEPC-treated RNase free ddH<sub>2</sub>O.
- Change gloves frequently and keep tubes closed.
- Reduce the preparation time as much as possible.
- **Use only sterile, disposable polypropylene tubes** throughout the procedure. (These tubes are generally RNase free.)
- Keep isolated RNA on ice.

This kit should only be used by personnel trained in *in vitro* diagnostic laboratory practice

### Storage of RNA

Purified RNA can be stored –80°C and is stable for months and years e.g. precipitated and stored in 70% ethanol.

### Quantification

Quantification of DNA and RNA from this assay must be done by means of qPCR or Reverse Transcriptase qPCR. All other methods will be disturbed by the included Carrier Nucleic Acids as well as DNA or RNA which is co-purified.

## Ordering information

Product	Package size	Catalogue No.
RTP® DNA/ RNA Virus Mini Kit	5 preparations	1040100100
RTP® DNA/ RNA Virus Mini Kit	50 preparations	1040100200
RTP® DNA/ RNA Virus Mini Kit	250 preparations	1040100300

### Single components for RTP® DNA/ RNA Virus Mini Kit

Extraction Tube (containing solid formula of lysis reagents incl. DNA control)	10 tubes	1040101500
Wash Buffer R1 (add 20 ml Ethanol)	20 ml	1040103500
Wash Buffer R2 (add 48 ml Ethanol)	12 ml	1040103600
Elution Buffer R	15 ml	1040104100

### Other sample sizes

Invisorb® Virus RNA HTS 96 Kit/ X	4 x 96 preparations	7143310300
Invisorb® Virus RNA HTS 96 Kit/ X	24 x 96 preparations	7143310400
Invisorb® Virus DNA HTS 96 Kit/ X	4 x 96 preparations	7142310300
Invisorb® Virus DNA HTS 96 Kit/ X	24 x 96 preparations	7142310400
InviMag® Virus RNA Kit/ KF96	1 x 96 preparations	7443300100
InviMag® Virus RNA Kit/ KF96	5 x 96 preparations	7443300200
InviMag® Virus DNA Kit/ KF96	1 x 96 preparations	7442300100
InviMag® Virus DNA Kit/ KF96	5 x 96 preparations	7442300200
InviMag® Virus DNA/ RNA Mini Kit	50 preparations	1440100200
InviMag® Virus DNA/ RNA Mini Kit	250 preparations	1440100300
InviMag® Virus DNA/RNA Mini Kit/ KFmL	15 preparations	2441150100
InviMag® Virus DNA/RNA Mini Kit/ KFmL	75 preparations	2441150200
Invisorb® Spin Virus DNA Mini Kit	50 preparations	1040200200
Invisorb® Spin Virus DNA Mini Kit	250 preparations	1040200300

### Possible suppliers for Isopropanol:

<b>Carl Roth</b> 2-Propanol Rotipuran >99.7%, p.a., ACS, ISO Ordering No. 6752	<b>Applichem</b> 2-Propanol für die Molekularbiologie Ordering No. A3928	<b>Sigma</b> 2-Propanol Ordering No. 59304-1L-F
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### Possible suppliers for Centrifuges:

<b>Eppendorf AG</b> 22331 Hamburg, Germany Phone: +49 (0) 40 53801 0 Fax: +49 (0) 40 53801 556 E-Mail: eppendorf@eppendorf.com Internet: www.eppendorf.com	<b>SIGMA Laborzentrifugen GmbH</b> 37507 Osterode am Harz, Germany Phone: +49-5522-5007-0 Fax: +49-5522-5007-12 E-Mail: info@sigma-zentrifugen.de Internet: www.sigma-zentrifugen.de
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